

Chapter 13

Lyophilization of microbial cultures: Principles, techniques, and applications in long-term preservation

Keshawanand Tripathi¹, Yashdeep Srivastava¹, Narendra Kumar^{2*}

¹ *Department of Biotechnology, Invertis University, Bareilly, Uttar Pradesh, India.*

² *School of Biotechnology and Bioengineering, Institute of Advanced Research, Gandhinagar, Gujarat, India.*

*Email: nkrathore1@gmail.com

1. Introduction:

Freeze drying (lyophilization), also known as lyophilization, is a widely used method for preserving microbial cultures by removing water from the samples while maintaining the integrity of the microbial cells (Atlas, 2010). This technique is particularly valuable for long-term storage of microorganisms, as it allows for the preservation of cell viability and functionality over extended periods (Smith, 2014; Tripathi et al., 2018; Yadav et al., 2021).

2. Principle:

2.1. Water Removal: Freeze drying involves freezing the microbial culture and then subjecting it to a vacuum, which allows the frozen water to sublime directly from the solid phase to the gas phase, bypassing the liquid phase. This process removes water from the microbial cells without causing significant damage to their structure.

2.2. Preservation of Cell Integrity: By removing water under low temperature and pressure conditions, freeze drying minimizes the formation of ice crystals within the microbial cells, which can cause cellular damage (Tiwari et al., 2023). This preservation method helps maintain the integrity and viability of the microbial cells for long-term storage.

2.3. Inactivation of Microbial Growth: The absence of water in the freeze-dried samples creates an inhospitable environment for microbial growth, effectively inhibiting the proliferation of contaminating microorganisms during storage.

2.4. Ease of Rehydration: Freeze-dried microbial cultures can be easily rehydrated by adding a suitable liquid medium, allowing the cells to return to their active state. This rehydration process enables the revival of preserved cultures for subsequent experimentation or culture maintenance.

3. Materials Required:

1. Pure culture of the microorganism
2. Sterile cryoprotective solution (e.g., glycerol or sucrose solution)
3. Sterile cryovials or ampules
4. Freeze dryer apparatus
5. Vacuum pump
6. Desiccant or other drying agents
7. Sterile pipettes and tips
8. Sterile distilled water
9. Refrigerator or freezer for storage

4. Procedure (Cappuccino and Welsh, 2019):

1. Start with a well-isolated and actively growing pure culture of the microorganism of interest. Ensure that the culture is in a healthy and vigorous state before preservation.
2. Prepare a cryoprotective solution by dissolving an appropriate cryoprotectant (e.g., glycerol or sucrose) in sterile distilled water to achieve a desired concentration (commonly 10-20%).
3. Mix the microbial culture with the cryoprotective solution in a sterile container (e.g., cryovial or ampule). Ensure thorough mixing to protect the microbial cells during the freezing and drying process.
4. Dispense the cryoprotected culture into sterile freeze-drying vials or ampules using sterile pipettes. Leave adequate headspace to accommodate expansion during freezing.
5. Place the filled vials or ampules in a laboratory freezer set to a temperature below the freezing point of water (typically -20°C to -80°C) and allow them to pre-freeze until solid.
6. Transfer the pre-frozen vials or ampules into the freeze dryer apparatus. Ensure that the samples are evenly spaced and securely positioned within the freeze-drying chamber.
7. Start the freeze-drying process by activating the vacuum pump to create a vacuum within the freeze-drying chamber. Gradually reduce the temperature within the chamber to induce sublimation of the frozen water from the samples.
8. Monitor the freeze-drying process closely, observing the temperature and pressure within the chamber. Adjust parameters as necessary to optimize the drying efficiency while minimizing damage to the microbial cells.
9. Allow the freeze-drying process to continue until all visible moisture has been removed from the samples. This typically takes several hours to overnight, depending on the size and composition of the samples.
10. Once the freeze-drying process is complete, seal the vials or ampoules under vacuum or with inert gas to prevent moisture ingress and microbial contamination.

11. Store the sealed freeze-dried samples in a refrigerator or freezer at a suitable temperature (-20°C to -80°C) for long-term preservation.

5. Precautions:

1. Ensure the microbial culture is in its exponential growth phase before starting.
2. Use sterile equipment and materials throughout the process.
3. Prepare a suitable cryoprotectant solution to protect the microbes during freeze-drying.
4. Label containers clearly with the culture name, date, and any other relevant information.
5. Freeze the culture rapidly to minimize damage from ice crystal formation.
6. Adjust the freezing rate and temperature according to the specific microbial strain.
7. Monitor and maintain consistent vacuum pressure during the drying process.
8. Avoid over-drying to prevent damage to the microbial cells.
9. Store the dried culture in sealed, moisture-proof containers at appropriate temperatures.
10. Validate the viability and stability of the preserved culture through periodic testing.

6. References:

1. Tiwari, P., Srivastava, Y., Sharma, A., & Vinayagam, R. (2023). Antimicrobial peptides: the production of novel peptide-based therapeutics in plant systems. *Life*, 13(9), 1875.
2. Atlas, R. M. (2010). *Handbook of microbiological media* (4th ed.). CRC Press.
3. Cappuccino, J. G., & Welsh, C. T. (2019). *Microbiology: A laboratory manual* (12th ed.). Pearson.
4. Tripathi, K., Kumar, N., & Abraham, G. (Eds.). (2018). *The role of photosynthetic microbes in agriculture and industry*. Nova Science Publishers, USA
5. Yadav, R. K., Chatrath, A., Tripathi, K., Gerard, M., Ahmad, A., Mishra, V., & Abraham, G. (2021). Salinity tolerance mechanism in the aquatic nitrogen fixing pteridophyte *Azolla*: a review. *Symbiosis*, 83, 129-142.
6. Tripathi, K., Kumar, N., Singh, M., & Singh, R. K. (2021). Fungal Siderophore: Biosynthesis, Transport, Regulation, and Potential. *Rhizosphere Microbes: Soil and Plant Functions*, 23, 387.